Staining of Blood Smears for Demonstration of Microfilariae and Haemoprotozoan Parasites



(Practical)



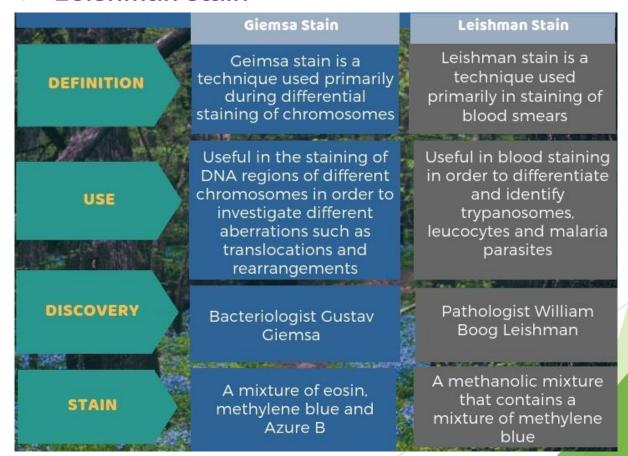
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Usually two types stains used in blood smears staining:-

- Giemsa stain
- Leishman stain



Leishman's stain:

This stain has a poor keeping quality in hot climate, so it is better to prepare a fresh stain from the powder every month or so.

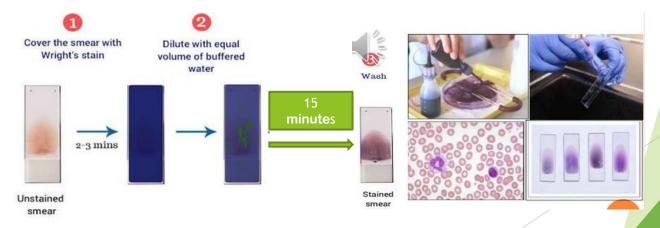
Leishman stain

- Composition
- Leishman powder (eosin-methylene blue powder) 0.5gm
- Acetone free methyl alcohol 100ml



Leishman stain:

- Prepare blood smear and air dry it.
- Pour Leishman stain dropwise (counting the drops) on the blood smear and allowed to act for 1- 2 minutes.
- Add double quantity of distilled water dropwise over the blood smears (i.e. double the numbers of dops).



Leishman's stain:

- Mixed by rocking the slide gently.
- 15 minutes are allowed for staining
- Slide is washed by stream of distilled water in horizontal position until the smear looks pink.
- o Thereafter, the slide is kept in a vertical position so that it drains and dries.

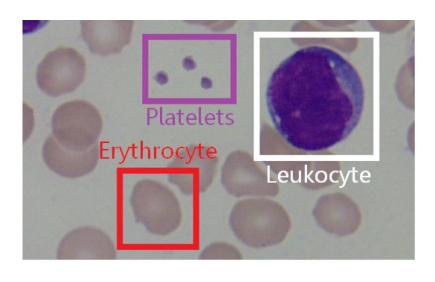
Leishman's stain:

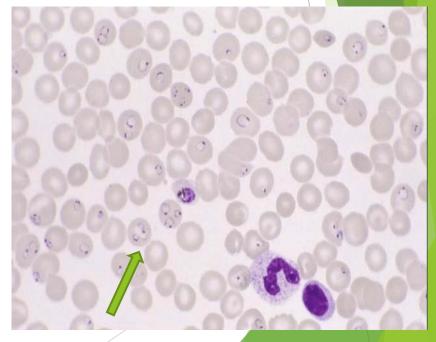
- Dry in air and examine under oil immersion lens of microscope.
- Fixation is not required in leishman's staining procedure because methyl alcohol is already mixed in leisman's stain.

Leishman's stain:

Microscopic examination.







Giemsa stain:

- Prepare blood smear and air dry it.
- Fixation: Fixed in methyl alcohol for two minutes
- Take out and dry it
- Pour the diluted Giemsa stain (1 in 10 dilutions) over the fixed smear
- (1 part Giemsa stain + 9 parts distilled water or buffer water)



Giemsa stain:

- It is kept for 45 minutes for allowing stains.
- Wash the smear under distilled water or tape water till the smear is just pink.
- Stained blood smears are dried in the air and examined under the microscope under oil immersion.



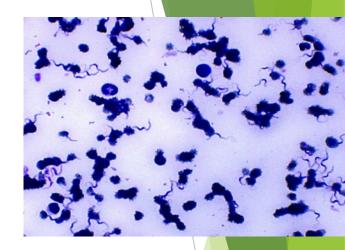


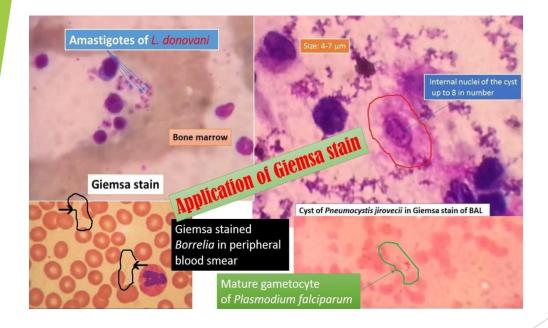
Giemsa stain:

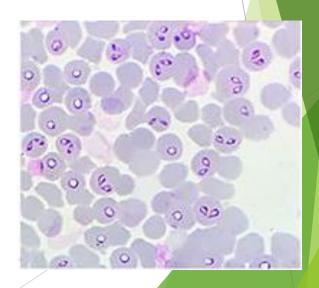
Microscopic examination:



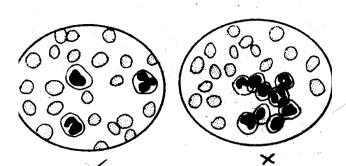








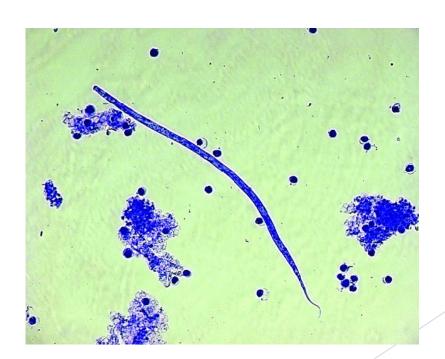
- Artifacts are liable to camouflage the precision of the microscopic findings.
- To avoid this, the slides should be placed vertically in the couplin jar having the required stain or preferably stained on a horizontal rack, diluted and washed with a buffer in the same position.
- If the stain is poured off, the scum is liable to stick to the slide and forms artifacts.



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Knott's Technique:-

It is used for the diagnosis of microfilariae
 of Dirofilaria immitis and Dipetalonema
 spp. through lyses of the red blood cells.



Knott's Technique:-

Materials:

- Suspected heparinized blood
- 2 % formalin
- Methylene blue
- Glass slide
- Coverslip
- Centrifuge tube
- Pipette

Knott's Technique:-

Procedure:

- Collect blood from the suspected dog in heparinized vial.
- Mix 1 ml of heparinized blood and 3 ml of 2% formalin
- Centrifuge the formalin mixed blood at 1500 rpm for 5-8 minutes.
- Discard the supernatant and then sediment is stained by adding equal amount of 1:1000 methylene blue stain.

Knott's Technique:-

Procedure:

- > One drop of mixture is placed on glass slide with the help of pipette and apply coverslip over it.
- Examine microfilariae under 40x or 100x

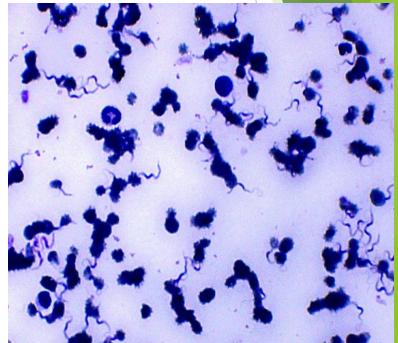




Microhaematocrit centrifuge technique

It is used for the detection of microfilariae of filarial worms or *Trypanosoma* spp. parasite.





Microhaematocrit centrifuge technique:

Materials:

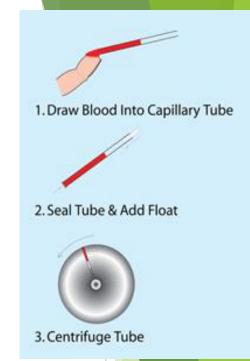
- Suspected blood samples
- Microhaematocrit tube
- Sealing materials or wax
- Microscope

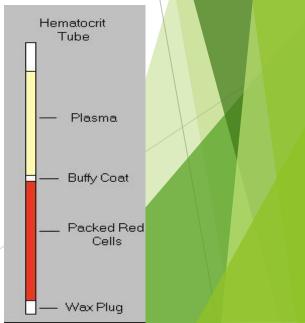


Blood smear preparation Microhaematocrit centrifuge technique:

Procedure-

- After filling the microhaematocrit tube with blood, centrifuge it for one minute.
- When suspected blood contained in microhaematocrit tube is centrifuged, three layers i.e. plasma layer, WBC or buffy coat layer and RBC layer.



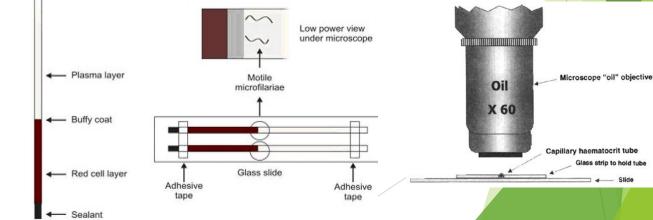


Microhaematocrit centrifuge technique:-

Procedure-

Then examine at the interface of the plasma and buffy coat for mortality. In the plasma microfilariae will be observed in active swimming condition

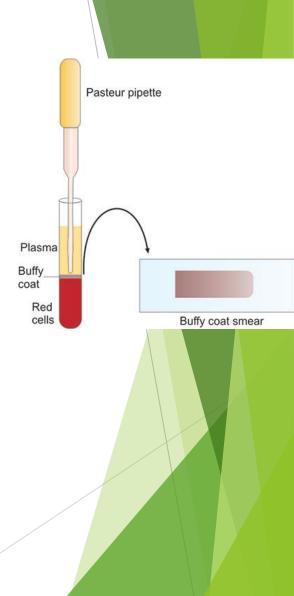
Microhematocrit tube after centrifugation



Microhaematocrit centrifuge technique:-

Procedure-

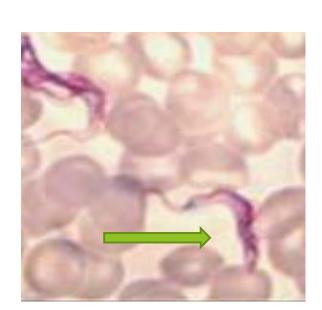
At the buffy coat interface break the tube and tab the buffy coat and a small amount of plasma on a slide; after placing a cover glass live microfilariae/ Trypanosoma spp. can be examined or allow it to air dry, fix for one minute with methyl alcohol and stain with Giemsa for 30 minutes for identification.



Microhaematocrit centrifuge

technique:-

Results: -Trypanosoma spp. and microfilaria may be observed in buffy coat smear after microscopic examination.





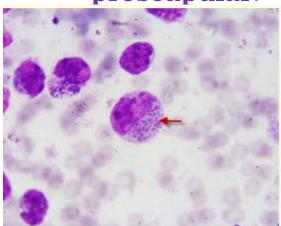
Plasma

Buffy

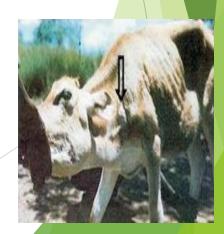
Pasteur pipette

Lymph Gland Biopsy:

- A lymph node smear is sometimes used in preference to a blood smear, for example during an investigation of theileiriosis or even trypanosomosis.
- Common lymph node used for biopsy is prescapular.







Kochs blue bodies

Lymph Gland Biopsy

Materials:

- Glass slide
- Shaving blade
- Methylated sprit
- Needle (16-18 gauze for cattle)

Enlargement of prescapular lymph nodes



Lymph Gland Biopsy: -

Procedure:

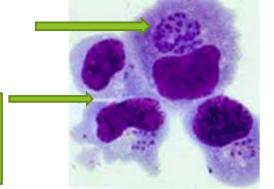
- > A suitable superficial lymph node is selected and palpated so that the site is fully known
- > Selected site is shaved with shaving blade and make antiseptic with the help of spirit
- > Punctured the palpated lymph node by a sterile hypodermic needle and lymph is gradually drawn into a clean syringe.
- Collected lymph fluids are ejected on a clean glass slide and make lymph smear with the help of a spreader slide
- > Then stain the lymph smear either with Giemsa or Leishman stain and examine under oil immersion lens of microscope.



Lymph Gland Biopsy:-

Result:

Schizont stages (Koch's blue bodies) of *Theileria* species found inside the cytoplasm of lymphocyte or Leaf like *Trypanosoma* sp. may be observed in lymph fluids.







THANK U